



# **Development of a Twin-Column Continuous Chromatography Oligo Purification Process** J Preston, PhD

Introduction

Peptide and Oligo synthesis technologies are well developed for large scale manufacturing of pharmaceutical materials. Isolation and purification is a critical step in these manufacturing processes, and almost always utilizes chromatography. Multi-column countercurrent solvent gradient purification (MCSGP) is a twin column continuous chromatography process that has significantly more capability than a traditional single-step batch process. The product yield is greatly improved with MCSGP while avoiding many complications encountered with a batch process.



#### Step 3: Run MCSGP

With MCSGP, a "cycle" is defined as both columns completing a gradient run. A "switch" is when a single column completes a gradient run.

It is best to run at least 4 cycles (8 switches) to evaluate the MCSGP methodology has reached steady state. The data can be viewed linearly as displayed below. The black trace below is the elution gradient. The red trace is the UV signal from column 1. The blue trace is the UV signal from column 2.



### MCSGP Technique Overview

**Typical Batch Process** 

pure product

late impurities are

later reprocessing

A typical batch process collects the pure portion of the desired peak. The portions with early and discarded or retained for purification continues

more pure product An MCSGP process collects the pure portion of the desired peak. The portions with early and late impurities are sent to a second column while adding more feed material and the

MCSGP Process

lecycle impur

# 2-Column MCSGP Process



The left column running a sample while the right column is being cleaned. Recycling the impure early eluting portion. Collecting the pure portion and adding more feed to the second column. Recycling the impure late eluting portion. The next step would be the right column running a sample while the left column is being cleaned.

The peak area data is entered into a spread sheet. The % purity of each fraction is calculated. The % yield of each fraction is also calculated.

Fraction	time							fraction	fractior
#	(min)	Imp A	Imp B	Imp C	Imp D	lmp E	Desired	% purity	% yield
4	34	0.0	0.0	0.0	0.0	0.0	85.4	100	0.1
5	35	0.0	0.0	152.8	140.9	0.0	1473.4	83	1.8
6	36	0.0	0.0	447.6	439.0	0.0	5438.0	86	6.5
7	37	0.0	158.0	656.6	514.7	0.0	9600.0	88	11.5
8	38	0.0	517.6	532.9	584.2	0.0	12298.0	88	14.7
9	39	0.0	857.7	558.9	721.0	0.0	11834.0	85	14.1
10	40	141.6	1133.4	251.0	296.3	339.2	9671.4	82	11.6
11	41	175.5	2687.9	252.8	265.4	617.6	7855.0	66	9.4
12	42	200.7	2562.1	294.4	275.0	1313.9	6319.7	58	7.5
13	43	133.7	2300.0	162.0	240.4	1604.0	5259.1	54	6.3
14	44	198.8	2139.0	212.5	280.1	2113.0	4371.9	47	5.2
15	45	0.0	1500.0	0.0	0.0	1825.1	3372.9	50	4.0
16	46	0.0	1300.0	0.0	0.0	1831.4	2517.3	45	3.0
17	47	0.0	1100.0	0.0	0.0	1339.9	1549.9	39	1.9
18	48	0.0	1078.0	0.0	0.0	1049.7	1069.7	33	1.3
19	49	0.0	770.8	0.0	0.0	1206.3	669.8	25	0.8
20	50	0.0	650.5	0.0	0.0	999.7	245.2	13	0.3
21	51	0.0	538.9	0.0	0.0	882.4	85.0	6	0.1
<u> </u>		0.0	000.0	0.0	0.0	002.4			0.1

To visualize the location of impurities under the desired peak. The peak areas for each peak are plotted vs the retention time for that peak.



## **Results and Discussion**

Analytical chromatogram and integration results for material collected during MCSGP experiment third cycle described above in Step 3. The results were similar for the second, third and fourth cycles. The results for the first cycle were slightly different due to the steady state not being reached yet.





TWIN **Production Skid** 



Available as 37 or 100 mL/min maximum flow rate. Capable of running MCSGP, single column batch, N-Rich, CaptureSMB

Custom built production equipment with flowrates up to 40 L/min. Capable of running MCSGP, single column batch, and separately – CaptureSMB.



0	2	4	6	8	10	12	14	m
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0						/,		<u> </u>
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With data from the Batch Fraction Analysis Table, expected purity and yield estimates can be made for different batch methodology pools. Purity estimates can also be made for MCSGP methodology based on this batch method data.

Batch		
Pool	% purity	% yield
Fr8	88.3	14.7
Fr7-Fr8	88.1	26.2
Fr6-Fr8	87.7	32.7
Fr6-Fr9	86.7	46.8
Fr6-Fr10	85.6	58.3
Fr6-Fr11	82.4	67.7

Fraction Analysis MCSGP results

By comparing the peak visualization data and the MCSGP Results, the collected material from the MCSGP is similar to the Fr6-Fr10 Batch Pool.

vield from MCSGP was 110%	
% yield obtained with the	Ba
Pool while the % purity for	Fr6
echnique was essentially	МС
cal.	

	% purity	% yield
atch Pool r6-Fr10	85.6	58.3
ICSGP Result	85.7	64.4

## Conclusions

MCSGP can be a valuable tool for oligo and peptide purifications. The twin-column technology allows for continuous recycling of the material that has insufficient purity. This allows for increased yields without added additional processing. The development of MCSGP methodology starts the same as developing batch methodology but results are then used by the MCSGP Wizard to generate the MCSGP methodology.

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Load Wizard File Current Wizard File	Exit		
Load Wizard File Save Wizard File Contectronifyizard folder/amgentmessp-04-21-2022.mcc The Wizard calculates the total amount of sample	Exit	Buffer Consumption Main Method   Pump P1 (mL) Buffer assiged to P1   465.1 /K. K. 20mM Sodium Phosphate dibasic,   225.6 B: B: 1M Sodium Phosphate dibasic,   Pump P2 [mL] Buffer assiged to P2   238.7 T: B: 1M Sodium Phosphate dibasic,   160.8 2: A 20mM Sodium Phosphate dibasic,   0 4: A 20mM Sodium Phosphate dibasic,   0 5: A 20mM Sodium Phosphate dibasic,	Overall Buff Pump P1 (r 523.3 263.7 Pump P2 (r 206.8 100.9 0 0 0
feed, buffers and eluents used for the MCSGP method and for the associated startup and shutdown methods.		0 6: A 20mM Sodium Phosphate dibasic,   0 7;   0 8:   Pump P3 (mL) Buffer assiged to P3   1 1: 1: 1: 1: 1: 0: 50dium   0 2: A 20mM Sodium Phosphate dibasic,   0 3: A 20mM Sodium Phosphate dibasic,   0 4: A 20mM Sodium Phosphate dibasic,   0 6: A 20mM Sodium Phosphate dibasic,   10.1 7: - sample 44mg-mL   0 8:   Overall Buffer Consumption Including Feed (mL)   1133 1133	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0

10.7 Total: 1133 [mL]

Generate Repor

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ChromIQ - MCSGP Wizard with Dynamic Process Control

atogram Washing & Regeneration Dynamic Process Control Method Settings & Performance Buffer Volumes Batch Method MCSGP Method



ChromaCon A new dimension in purification

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